

FGFR antagonists restore defective mandibular bone repair in a mouse model of osteochondrodysplasia



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Background

Hypochondroplasia (HCH) is linked to Fibroblast Growth Factor Receptor 3 (FGFR3) gain-of-function mutations. These patients are characterized by rhizomelic dwarfism, and craniomaxillofacial anomalies requiring craniomaxillofacial bone surgeries. Our team generated the first Hch mouse model (Fgfr3^{N534K/+}) (Loisay et al, 2023), expressing the most common missense FGFR3 mutation (p.Asn540Lys), presenting progressive dwarfism, maxillary retrusion and mandibular prognathism mimicking human pathology.

Previously we demonstrated that mandibular bone healing was impaired in this Hch mouse model, we observed a defective bone consolidation, significant microarchitectural alterations of the calluses and abnormal presence of pseudarthrosis (Morice et al, Bone



Analyzing the effects of two FGFR3 antagonists on mandibular **bone repair** in this *Hch* mouse model:

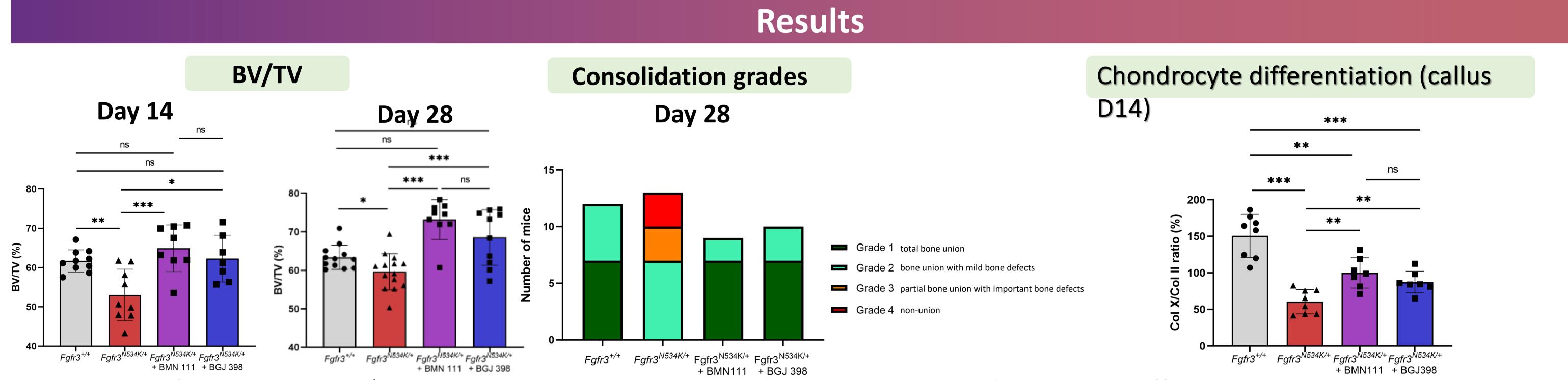


1) a tyrosine kinase inhibitor, Infigratinib (BGJ398) (phase II clinical trial for achondroplasia children); 2) vosoritide, a CNP analog (BMN 111) (Voxzogo[®]) with FDA and EC

approval for achondroplasia children.

We performed **unstabilized fractures** at 6 weeks of age on the right mandible in *Hch* mice. **DO** Fracture **D7** D21 **D28** Soft callus Bone callus

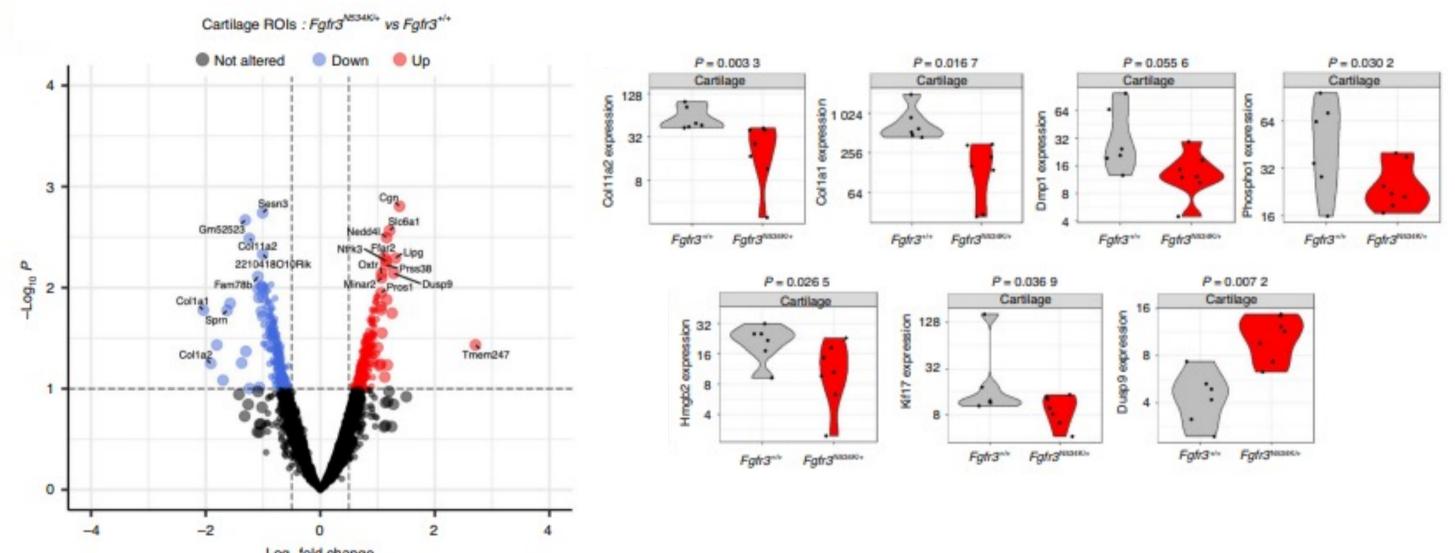
At day 0 post fracture, subcutaneous injections were performed with BGJ398 (4 mg/kg) or BMN111 (0.8 mg/kg) 3 times/week, during either 14 or 28 days. Morphometric analyses concerned two major key points of bone repair, D14 and D28 post fracture.

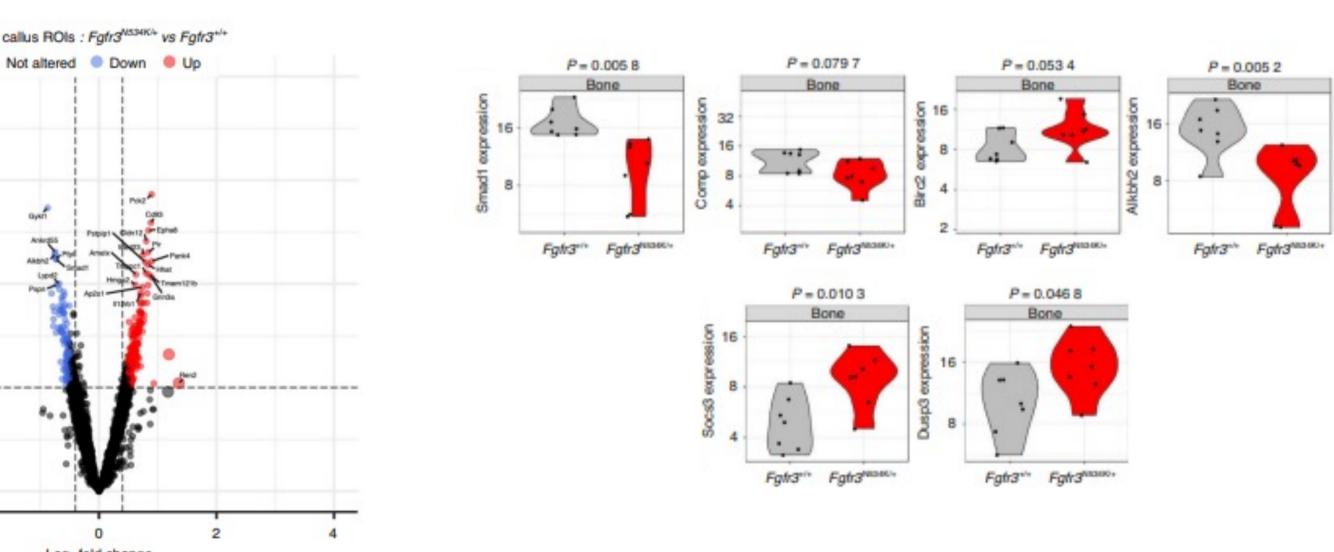


Bone Volume/Total Volume (BV/TV) values of the calluses were improved in treated groups: D14 (+29 %, p<0.005 (BMN 111), +20.8 %, p<0.05 (BGJ 398)); D28 (+ 24%, p<0.005 BMN 111), (+ 17,8%, p<0.01 BGJ 398). Calluses volumes were significantly reduced at D28: -30.9 %, p<0.005 (BMN 111); - 38,6 %, p<0.0001 (BGJ 398), corresponding to accelerated bone remodeling. All treated mutants presented normal grades of consolidation (grades 1 or 2), without pseudarthrosis (grade 4)

Chondrocyte differentiation was significantly improved by both treatments: significant increase of the ratio of Col X / CollI chondrocyte positive area : BMN111: +64.9 %, p<0.01; BGJ398: +40.6 %, p<0.01 in *Fgfr3^{N534K/+}* mice compared to control mice.

at D28. Spatial transcriptomic analysis of the cartilage and newly formed bone in the calluses from Fgfr3^{N534K/+} and Fgfr3^{+/+} mice at day 14 post-fracture





Volcano plot representations of differentially expressed genes (P<0.1) that were either upregulated (in red) or downregulated (in blue) in the cartilage of the calluses in Fqfr3^{N534K/+} mice, relative to $Fgfr3^{+/+}$ mice.

Violin plots showing the expression levels (normalized read counts) of Col11, Col1a1, Dmp1, Phospho1, Hmgb2, Kif17 and Dusp9 in the cartilage of the calluses from Fgfr3^{N534K/+} mice (in red) and Fgfr3^{+/+}

Volcano plot representations of differentially expressed genes (P<0.1) that were either upregulated (in red) or downregulated (in blue) in the bone of the calluses in *Fgfr3^{N534K/+}* mice, relative to *Fgfr3^{+/+}* mice. Violin plots showing the expression levels (normalized read counts) of Smad1, Comp, Birc2, Alkbh2, Dusp3 and Socs3 in the newly formed bone of the calluses from *Fgfr3^{N534K/+}* mice (in red) and *Fgfr3^{+/+}* mice (in grey).

Conclusions and perspectives

Our findings suggest that both MAPK and STAT pathways are over-activated in the bone callus in a mouse model of hypochondroplasia, and thus highlight the key role of these downstream signaling pathway in FGFR3-related disorders.

Our data showed that the constitutive activation of FGFR3 was counteracted by infigratinib and vosorotide thus restoring the defective mandibular endochondral bone repair, this modulation of FGFR3 signaling might be of value for treating bone repair in patients with FGFR3 chondrodysplasia.